

The preparation of hexasilsesquioxane (T₆) cages by “non aqueous” hydrolysis of trichlorosilanes

Alan R. Bassindale,^{*a} Iain A. MacKinnon,^b Maria G. Maesano^a and Peter G. Taylor^{*a}

^a The Chemistry Department, The Open University, Walton Hall, Milton Keynes, UK MK7 6AA.

E-mail: P.G.Taylor@open.ac.uk; Fax: (+44) 1908 858327; Tel: (+44) 1908 652512

^b Dow Corning Ltd, Cardiff Road, Barry, South Glamorgan, UK CF63 2YL

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Hexasilsesquioxane cages (T₆) have been prepared from a range of alkyl and aryl trichlorosilanes using a “non aqueous” hydrolysis with dimethyl sulfoxide.

T₆ silsesquioxane cages (RSiO_{1.5})₆ have received a great deal of attention as cores for building octopus molecules¹ and dendrimers² and as models of resins. Such cages have also been used for the preparation of partially opened cages that themselves act as models for silica surfaces and supports for transition metal catalysts.^{3–6} However, only a limited range of spherical organosilsesquioxanes have been prepared even though a large variety of trichlorosilanes are available.⁷ This is because conventional aqueous hydrolysis, including “scarce water hydrolysis” leads to mainly resin and a large variety of cage and open cage compounds. Only with specific alkyl groups is particular cage formation favoured to such an extent and/or solubility differences large enough to enable isolation of pure silsesquioxane cages. Nevertheless the yields obtained by these methods are very poor and the octasilsesquioxanes, (RSiO_{1.5})₈, T₈, seem to be the preferred outcome. An alternative route to such species is the formation of the spherical hydrogen silsesquioxane followed by hydrosilylation using alkene.¹ We have used this approach to build T₈ molecules with a range of functionalities such as dendrimers, liquid crystals and surfactants. We are interested in the preparation of functionalised hexasilsesquioxanes, T₆, and the mechanism of their conversion into T₈ and larger cage structures.⁸ However, we have been unable to prepare hydrogen hexasilsesquioxane for use in our hydrosilylation methodology and therefore we examined alternative methods of T₆ formation.

Hexa(cyclohexylsilsesquioxane) has been prepared by Feher³ using a modification of Brown’s method⁹, involving aqueous hydrolysis in a water/acetone mixture. Reasonable yields are obtained, but only after long periods. For example, after four months of hydrolysis, Molloy obtained sufficient crystals to carry out an X-ray crystal structure analysis.¹⁰ We have obtained reasonable yields of this T₆ molecule in a shorter time using a similar aqueous/organic hydrolysis, however, this route is limited to a few compact substituents with secondary or tertiary groups adjacent to the silicon. Matsumoto has developed an alternative approach to T₆ based on the dicyclohexylcarbodiimide coupling of the silane triol or 1,1,3,3-tetrahydroxydisiloxane.¹¹ Unfortunately this route seems only possible with bulky substituents such as *tert*-butyl or 1,1,2-trimethylpropyl.

We thus looked for an alternative method of constructing the T₆ cage that was easy to carry out, used readily available starting materials and was applicable to a wide range of substituents on silicon. The “non-aqueous” hydrolysis of dichlorosilane using dimethyl sulfoxide was first reported in

1965¹² and has subsequently been revisited by Voronkov¹³, Weber¹⁴ and Brook.¹⁵

Brook’s most recent study has shown that early in the reaction the six membered cyclotrisiloxane, D₃, is formed in good yields. If six membered rings are preferred with trichlorosilanes this should favour T₆ formation through the ring acting as a template or through the coupling of two such T₃ rings.

Table 1 gives the results of our study†. We found that trichlorosilanes could be reacted with dimethyl sulfoxide (2 equivalents) in chloroform at room temperature to give T₆

Table 1 Yields and properties of T₆ cages prepared by the “non aqueous” hydrolysis of trichlorosilanes using DMSO

Starting trichlorosilane	²⁹ Si-NMR/ppm	Physical state of T ₆ product	Yield/%
Octyltrichlorosilane	–54.2	Gel	25
Cyclohexyltrichlorosilane	–56.6	Solid	11
Cyclopentyltrichlorosilane	–54.4	Solid	9
2-Methylpropyltrichlorosilane	–55.4	Solid	11
Phenyltrichlorosilane	–66.9	Solid	7
3-(<i>p</i> -Methoxyphenyl)propyl trichlorosilane	–54.4	Solid	6

cages. All yields quoted are isolated yields after purification. The T₆ cage is formed together with resin by-products, which are sometimes difficult to separate. Silsesquioxane cages are often purified by column chromatography, however, because of the T₃ rings we found that the T₆ cages were particularly susceptible to decomposition in the presence of nucleophilic species. Thus the T₆ cages decomposed on the silica or alumina stationary phases used in column chromatography. However, we found that treating the silica gel with 5–10% (by mass) trimethylsilyl chloride before elution of the reaction mixture gave an ideal chromatography medium that was sufficiently active to maintain separation but not too active to cause degradation of the T₆. Although the yields are low in absolute terms, they are perfectly acceptable when compared to conventional cage syntheses. For example, scarce water hydrolysis of trichlorosilanes gives T₈ cages in about 15% yield. We have carried out the “non aqueous” hydrolysis of dichlorosilanes to give D₃, D₄, D₅ and D₆ rings using a range of other high oxidation state oxygen donors such as triphenylphosphine oxide, pyridine *N*-oxide, iodosobenzene and sulfur trioxide/pyridine. However, DMSO was the only reagent that gave specific cage compounds with organotrichlorosilanes.

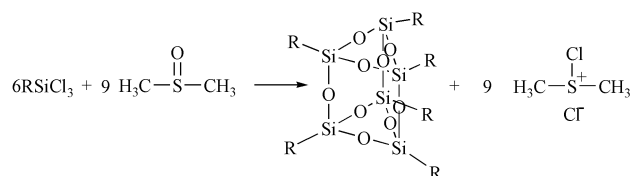
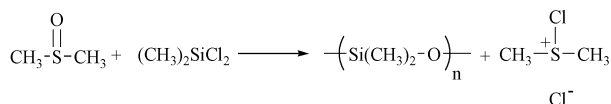
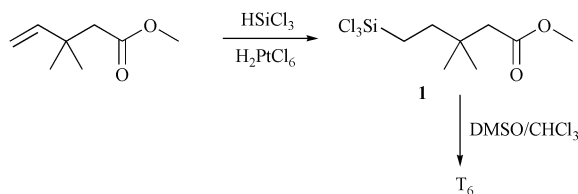


Table 1 shows that, as expected, compact substituents such as cyclopentyl, cyclohexyl and phenyl give T_6 cages in acceptable yields. Surprisingly, primary alkyltrichlorosilanes such as octyl, 2-methylpropyl and 3-(*p*-methoxyphenyl)propyl gave T_6 cages in comparatively good yields and this represents the first synthesis of this important class of compounds. In contrast to the Matsumoto route, we were unable to isolate T_6 cages when bulky substituents such as *tert*-butyl were employed. In this case a range of products were observed which were probably D polyol rings. Similarly cage structures were not produced with small substituents, for example, vinyltrichlorosilane gave no identifiable products. Chan has shown that HSiCl_3 reduces DMSO to dimethyl sulfide.¹⁶ In contrast we have been able to form D_3 and D_4 rings from $\text{CH}_3\text{SiHCl}_2$ using DMSO, however, reaction of DMSO with trichlorosilane led to a complex mixture of products.

Our previous strategy for preparing organooctasilsesquioxanes relied on the synthesis of the alkene arm followed by attachment to hydrogen silsesquioxane by hydrosilylation.¹ We have been able to transfer this methodology to T_6 synthesis through hydrosilylation of the alkene with trichlorosilane followed by reaction with DMSO. For example, hydrosilylation of methyl 3,3-dimethylpent-4-enoate using chloroplatinic acid gives the corresponding trichlorosilane derivative, **1**, in 77% yield and thence the T_6 derivative in 6.3% yield.



As with the T_8 cages we are interested in preparing polyfunctional cages and dendrimers. This requires the synthesis of cages with functionality on the arms. Reaction of the methyl dimethylpentanoate derivative with ethanediamine followed by methyl acrylate will give a dendrimer with twelve ester groups.¹⁷

The ^{29}Si NMR chemical shifts of the T_6 cage silicons are usually in the region -54 ppm to -57 ppm. This is in agreement with the chemical shifts of previously prepared T_6 cages.^{4,11} The exception to this is phenyl hexasilsesquioxane which has a ^{29}Si NMR chemical shift of -66.89 ppm. This upfield shift is typical of a phenyl substituent, as observed in the D ring series. The corresponding range for T_8 cages is -65 ppm to -67 ppm. Marsmann and co-workers have developed a method to estimate the ^{29}Si NMR chemical shift of an unknown T_{10} or T_{12} cage based on the chemical shift of the corresponding T_8 cage.¹⁸ For example, for T_{10} cages the equation $\delta_{T_{10}} = 1.028 \times \delta_{T_8}$ holds.

A plot of the ^{29}Si NMR chemical shift of the T_6 cages in Table 1 versus the chemical shift of the corresponding T_8 cages has a slope of 0.82 ($R^2 = 0.94$ (forced to go through the origin)), suggesting the equation $\delta_{T_6} = 0.82 \times \delta_{T_8}$.

The chemical shifts estimated using this equation correlate well with the measured chemical shifts in Table 1, however, differ most significantly for the phenyl and benzyl substituents. The size of the constant in the equations above reflects the size of the rings associated with the silicon in question. A silicon in a T_8 is associated with three T_4 rings, whereas a silicon in a T_{10} is associated with two T_4 rings and a T_5 ring, the relief of angle strain leading to a slightly higher factor. A silicon in a T_6 cage is associated with one T_3 ring and two T_4 rings, the increase in ring strain leading to lower factor and the decrease is relatively large. This matches the behaviour in D_5 , D_4 and D_3 rings.

In conclusion we have developed a robust method for the synthesis of T_6 cages in reasonable yield with a wide range of substituents. This should lead to T_6 structures being used in exactly the same way that T_8 cages are employed for building octopus molecules and dendrimers, as models of resins and as

starting materials for the preparation of partially opened cages.

Notes and references

† Preparation of T_6 cages: A solution of DMSO (4 mL, 54 mmol) in chloroform (35 mL) was added to a solution of the trichlorosilane (27 mmol) in chloroform (40 mL) and the mixture stirred at room temperature for 24 h. The mixture was washed with water (4×20 mL), dried with MgSO_4 and the solvent removed under reduced pressure. Column chromatography of the resulting gel using chloroform as the eluant gave a pure sample of the T_6 cage compound.

Hexa(octylsilsesquioxane); yield 25%; δ_{H} (300 MHz, CDCl_3 , Me_4Si) 0.10 (2H, br, $\text{CH}_2\text{-Si}$), 0.36 (3H, t, CH_3), 0.74 (12H, br, CH_2); δ_{C} (75 MHz, CDCl_3 , Me_4Si) 11.36 ($\text{CH}_2\text{-Si}$), 14.07 (CH_3), 22.28 (CH_2), 22.65 (CH_2), 29.23 (CH_2), 30.85 (CH_2), 31.91 (CH_2), 32.54 (CH_2); δ_{Si} (79.3 MHz, CDCl_3 , Me_4Si) -54.2 ; m/z (MALDI) 1013 [$\text{M} + \text{Na}^+$]; EA: Found: C, 58.10; H, 10.51. $\text{C}_{48}\text{H}_{102}\text{Si}_6\text{O}_9$ requires C, 58.18; H, 10.30%.

Hexa(cyclohexylsilsesquioxane)¹⁰; yield 11%; δ_{H} (300 MHz, CDCl_3 , Me_4Si) 0.78 (1H, br, CH-Si), 1.18 (5H, br, $\text{CH}_{(\text{ax})}$), 1.71 (5H, br, $\text{CH}_{(\text{eq})}$); δ_{C} (75 MHz, CDCl_3 , Me_4Si) 22.6 (CH), 26.1 (2 CH_2), 26.6 (2 CH_2), 27.2 (CH_2); δ_{Si} (79.3 MHz, CDCl_3 , Me_4Si) -56.6 ; m/z (EI) 811 [MH^+], 730.

Hexa(cyclopentylsilsesquioxane); yield 9%; δ_{H} (300 MHz, CDCl_3 , Me_4Si) 0.97 (1H, t, CH-Si), 1.45 (6H, br, CH_2), 1.71 (2H, br, CH_2); δ_{C} (75 MHz, CDCl_3 , Me_4Si) 21.68, 26.80, 26.97; δ_{Si} (79.3 MHz, CDCl_3 , Me_4Si) -54.4 ; m/z (EI) 726 [M^+]. This was successfully analysed by X-ray crystallography to confirm the structure. This will be discussed in detail in a subsequent publication.

Hexa(2-methylpropylsilsesquioxane); yield 11%; δ_{H} (300 MHz, CDCl_3 , Me_4Si) 0.63 (2H, d, CH_2), 0.96 (2H, d, 2 CH_3), 1.78 (1H, m, CH); δ_{C} (75 MHz, CDCl_3 , Me_4Si) 22.0, 23.8, 25.6; δ_{Si} (79.3 MHz, CDCl_3 , Me_4Si) -54.4 ; m/z (EI) 654 [M^+]; EA: Found: C, 44.25; H, 8.49. $\text{C}_{24}\text{H}_{54}\text{Si}_6\text{O}_9$ requires C, 44.00; H, 8.31%.

Hexa(phenylsilsesquioxane); yield 7%; δ_{H} (300 MHz, CDCl_3 , Me_4Si) 6.98 (5H, m, Ph) δ_{C} (75 MHz, CDCl_3 , Me_4Si) 127.5, 130.3, 131.8, 134.2; δ_{Si} (79.3 MHz, CDCl_3 , Me_4Si) -66.9 ; m/z (EI) 774 [M^+]; EA: Found: C, 55.16; H, 4.14. $\text{C}_{36}\text{H}_{30}\text{Si}_6\text{O}_9$ requires C, 55.88; H, 3.87%.

Hexa(3-(*p*-methoxyphenyl)propylsilsesquioxane); yield 6%; δ_{H} (300 MHz, CDCl_3 , Me_4Si) 0.65 (2H, t, CH_2), 1.68 (2H, p, CH_2), 2.53 (2H, t, CH_2), 3.73 (3H, s, CH_3), 6.7 (2H, d, Ph), 6.9 (2H, d, Ph); δ_{C} (75 MHz, CDCl_3 , Me_4Si) 11.0 ($\text{CH}_2\text{-Si}$), 24.4 (CH_2), 37.6 ($\text{CH}_2\text{-Ph}$), 55.3 (CH_3), 113.7 (Ph), 129.4 (Ph), 134.2 (Ph), 157.8 (Ph); δ_{Si} (79.3 MHz, CDCl_3 , Me_4Si) -54.4 ; m/z (EI) 1205 [M^+]. This was successfully analysed by X-ray crystallography to confirm the structure. This will be discussed in detail in a subsequent publication.

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